

The use of microcurrent bioelectric stimulation to reverse erectile dysfunction: A randomized, double-blind, placebo-controlled trial.

Background:

Erectile dysfunction (ED) affects up to 20% of men, and an increasing problem with advancing age. There are many contributing causes of ED, including concomitant use of prescribed medications for health problems including hypertension, or causal diseases such as diabetes. The current treatment includes improved diet, exercise, stress reduction and sleep, but also the use of drugs that are inhibitors of the Phosphodiesterase pathway, which leads to vasodilation in the venous system of the penis to induce erections. These drugs have significant long-term side effects, and are very expensive, prohibiting their use by a large percentage of men with ED.

There are a number of electrical fields in the body, with examples including the electrocardiogram (EKG) and brain encephalogram (EEG). Bioelectric stimulation (BES) utilizes the delivery of precise microcurrents targeted to specific proteins in the tissue being treated that causes upregulation or increased local tissue expression of the protein(s) of interest. The identification of the precise signal for each target protein has taken decades to define and confirm, using quantitative measurements from tissues that were stimulated at various frequencies until the optimal signal was identified. The use of BES has been shown to be very safe, and in fact is being tested as a treatment for cancer. It has been used to induce organ and tissue repair and regeneration in many diseases and conditions, varying from accelerating tooth movement, reversing heart failure, traumatic brain injury and stroke, to enhanced breast milk production, and hair regeneration.

This study follows the very successful demonstration of the absolute safety, and remarkable efficacy, of using functional bioelectric stimulation to treat ED by Carboni and colleagues¹. Dr Carboni will also serve as the Principal Investigator of this new follow up study. The previous study was a very well designed randomized, sham and placebo-controlled, trial that demonstrated 100 % success in reversing ED with no other intervention in the treatment cohort, while none of the control subjects experienced reversal of ED during the 4 week study period. The bioelectric signal used in the study targeted only a single protein, vascular

endothelial growth factor (VEGF), which has been shown to be one of the most potent inducers of the formation of new blood vessels and improve blood flow.

The goal of this study will be to examine the potentially additive effect of adding a third arm to the previous protocol to include the addition of 4 new target proteins to VEGF including endothelial nitric oxide (eNOS), which regulates endothelial vasoreactivity and should enhance venous vasodilation, stromal derived factor-1 (SDF-1) which stabilizes the blood vessels formed,), insulin-like growth factor (IGF) which is very decreased in ED, and follistatin, which is a potent stimulator of muscle function and contractility leading to enhanced erection, all leading to improved blood flow and reversal of ED.

Study Design: Randomized, double-blind, sham and placebo-controlled

Number of Study Subjects: 30, with 10 subjects in each of 3 groups

Number of Sites: one

Department of Health Science and Rehabilitation, Federal University of Health Sciences of Porto Alegre–UFCSPA, Porto Alegre, Rio Grande do Sul, Brazil

Principal Investigator: Cristiane Carboni, Msc, Physiotherapy

Study Enrollment: No subject will receive treatment until they have had all possible risks and benefits explained by the PI and the Consent Form signed.

Expected Duration of Enrollment: 3 months

Study Arms: 3

Gp I: Control. To receive the same duration of BES in a blinded manner as the other two groups, but no BES delivered.

Gp II: BES at the same current, duration, and frequency as the previous study, and other treatment groups, but targeted to only VEGF.

Gp III: BES of the same duration as all groups, but targeted to not only VEGF, but to 4 additional proteins (eNOS, SDF, IGF, and Follistatin).

Treatment:

Number of total treatments: 8

Frequency: twice/week for 4 weeks

Duration of treatment: 20 minutes/treatment

Stimulator to be used: Commercially approved stimulator, either the one used in the previous study conducted by Carboni et al¹ for this indication (Neurodyne), or

other commercial approved stimulator that is confirmed to be able to reach the signals required for the additional protein targets.

Method of Delivery of stimulation: Simple patch electrodes applied to the dorsum of the penis and connected to the bioelectric stimulator. The current will be slowly increased from zero to the specific current for each protein to be stimulated.

The stimulator will be controlled by a medical assistant who will prevent the subject from seeing any adjustment of current, with no presence of the PI or Co-PI's in order to maintain the double blind of the study design. Care will be taken to assure each subject receives the same duration of actual or pretend stimulation to maintain the blind for each subject in the study.

End Points:

1. Reversal of self-assessed ED compared to baseline on a scale of 1-5.
2. Adverse events including pain, local skin irritation, or difficulty with urination.
3. Serum Testosterone levels will be measured in 3 randomly selected subjects in each of the three treatment groups at baseline, one hour after a treatment following 2 weeks of study, and at the end of the study. These blood levels are to examine the potential role of BES in stimulating increased testosterone production, and a possible role in ED reversal.

Data Analysis:

Patients will be seen by the PI or sub-investigator at the time of study enrollment, the mid-point(2 weeks), and end of the study (4 weeks) to assess any adverse events. However each subject's self-assessment of the degree of reversal of ED will be collected by a person blinded to treatment assignment to prevent bias in data analysis.

References:

1. Carboni C, Fornari A, Bragante K, Averbeck A, Vianna da Rosa P, Della Mea Plentz R. An initial study on the effect of functional electrical stimulation in erectile dysfunction: a randomized controlled trial. *Internatl J Impotence*. 2018; 30:97-101
2. Otunctemur A, Ozbek E, Sahin S, Ozcan L, Dursun M, Polat EC, Cekmen M, Ozsoy OD, Erkoc M, Danis E, Bozkurt M. Low serum insulin-like growth factor-1 in patients with erectile dysfunction. *Basic Clin Androl*. 2016 Jan 28;26:1.
3. Xu ZP, Wang HP, Liu JM, Zheng XG, Wu D, Pu XY. Effects of insulin-like growth factor-1 on the relaxation responses of the cavernous smooth muscle from aged rats. *Scand J Urol*. 2015 Jun;49(3):260-6.
4. NIH Consensus Development Panel on Impotence. Impotence- NIH Consensus Conference. *JAMA*. 2013;270:83–90.
5. Pournaghash-Tehrani S, Etemadi S. ED and quality of life in CABG patients: an intervention study using PRECEDE- PROCEED educational program. *Int J Impot Res*. 2014;26:16–9.
6. McMahon CN, Smith CJ, Shabsigh R. Treating erectile dysfunction when PDE5 inhibitors fail. *BMJ*. 2006;332:589–92.
7. Al-Shaiji T, Brock G. Phosphodiesterase Type 5 inhibitors for the management of erectile dysfunction: preference and adherence to treatment. *Curr Pharm Des*. 2009;15:3486–95.
8. Hwancheol S, Kwanjin P, Soo-Woong K, Jae-Seung P. Reasons for discontinuation of sildenafil citrate after successful restoration of erectile function. *Asian J Androl*. 2004;6:117–20.
9. Claes H, Van Kampen M, Lysens R, Baert L. Pelvic floor exercise in the treatment of impotence. *Eur J Phys Med Rehabil*. 1995;5:42–6.

10. Derouet H, Nolden W, Jost W, Osterhage J, Eckert R, Ziegler M. Treatment of erectile dysfunction by an external ischiocavernous muscle stimulator. *Eur Urol.* 1998;34:355–9.
11. Andersson K-E Erectile physiological and pathophysiological pathways involved in erectile dysfunction. *J Urol.* 2003;170(2 Pt 2):S6-13-4.
12. Hurt KJ, Musicki B, Palese Ma, Crone JK, Becker RE, Moriarity JL, et al. Akt-dependent phosphorylation of endothelial nitric- oxide synthase mediates penile erection. *Proc Natl Acad Sci USA.* 2002;99:4061–6.
13. Gratzke C, Angulo J, Chitale K, Dai Y-T, Kim NN, Paick J-S, et al. Anatomy, physiology, and pathophysiology of erectile dysfunction. *J Sex Med.* 2010;7(1 Pt 2):445–75.
14. Stief CG, Weller E, Noack T, Djamilian M, Meschi M, Truss M, et al. Functional electromyostimulation of the corpus cavernosum penis--preliminary results of a novel therapeutic option for erectile dysfunction. *World J Urol.* 1995;13:243–7.
15. Dean R, Lue TF. Physiology of penile erection and pathophysiology of erectile dysfunction. *Urol Clin North Am.* 2005;32:379–95.
16. Ignarro LJ, Bush PA, Buga GM, Wood KS, Fukuto JM, Rajfer J. Nitric oxide and cyclic GMP formation upon electrical field stimulation cause relaxation of corpus cavernosum smooth muscle. *Biochem Biophys Res Commun.* 1990;170:843–50.
17. Jiang J, He Y, Jiang R. Ultrastructural changes of penile cavernous tissue in multiple sclerotic rats. *J Sex Med.* 2009;6: 2206–14.
18. Hatzimouratidis K, Amar E, Eardley A, Giuliano F, Hatzichristou D, Montorsi F, et al. Guidelines on male sexual dysfunction: erectile dysfunction and premature ejaculation. *Eur Urol.* 2010;57: 804–14.
19. Paick J, Goldsmith P, Barta A, Nunes L, Padula C, Lue T. Relationship between venous incompetence and cavernous nerve injury: Ultrastructural alteration of cavernous smooth muscle in the neurotomized dog. *Int J Impot Res.* 1991;3:173–84.

20. Myung-Cheol G, Yun-Chul O, Tae-Woo K. The effect of treatment of erectile dysfunction with electrical stimulation. *Kor J Androl*. 2000;18:149–55.
21. Althof S. Quality of life and erectile dysfunction. *Urology*. 2002;59:803–10.
22. Araujo A, Durante R, Feldman H, Goldstein I, McKinlay J. The relationship between depressive symptoms and male erectile dysfunction: cross-sectional results from the Mass. Male Aging Study. *Psychosom Med*. 1998;60:458–65.
23. Lewis RW, Fugl-Meyer KS, Corona G, Hayes RD, Laumann EO, Moreira ED, et al. Definitions/epidemiology/risk factors for sexual dysfunction. *J Sex Med*. 2010;7(4 Pt 2):1598–607.
24. Kanno S, Oda N, Abe M, Saito S, Hori K, Handa Y, Tabayashi K, Sato Y. Establishment of a Simple and Practical Procedure Applicable to Therapeutic Angiogenesis. *Circulation*. 1999;99(20):2682–7.
25. Hunckler J, de Mel A. A current affair: electrotherapy for wound healing. *J or Multidisciplinary Healthcare*. 2017; 10: 179-194.
26. Birmingham k, Gradinaru V, Ludwig K, Famm K. Bioelectronic medicines: A research roadmap. *Nat Rev Drug Discovery*. 2014;13(6):399-407
27. McLaughlin KA, Levin M. Bioelectric signaling in regeneration: Mechanisms of ionic controls of growth and form. *Dev Biol*. 2018 Jan 15;433(2):177-189
28. Baer ML, Colello RJ. Endogenous bioelectric fields: A putative regulator of repair and wound regeneration. *Neural Regen Res*. 2016 Jun;11(6):861-64.
29. Bassett CA. Beneficial effects of electromagnetic fields. *J Cell Biochem*. 1993 Apr;51(4):387-93
30. Roth BJ, Wikswo JP Jr. Electrically silent magnetic fields. *Biophys J*. 1986 Oct;50(4):739-45.
31. Hoare JI, Rajnicek AM, McCaig CD, Barker RN, Wilson HM. Electric fields are novel determinants of human macrophage functions. *J Leukocyte Biol*. 2016;99(6):1141–1151.

32. Peters EJ, Armstrong DG, Wunderlich RP, Bosma J, Stacpoole-Shea S, Lavery LA. The benefit of electrical stimulation to enhance perfusion in persons with diabetes mellitus. *The Journal of Foot and Ankle Surgery*. 1998;37(5):396–400.

33. Park RJ, Son H, Kim K, Kim S, Oh T. The Effect of Microcurrent Electrical Stimulation on the Blood Circulation of the Foot and Pain of Diabetic Neuropathy. *Journal of Physical Therapy Science*. 2011;23(3):515–8.